

Bölüm 37

İMMUNOTERAPİ İLE SİSTEMİK KEMOTERAPİNİN KOMBİNASYON TEDAVİLERİ

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GİRİŞ

İmmünoloji ve onkoloji alanları, cerrah William Coley'nin öldürülmüş bakterilerin sarkom bölgelerine enjeksiyonunun tümör küçülmesine yol açabileceğini bildirdiği 19. yüzyılın sonlarından beri gelişimleri birbirlerine bağlantılı bir şekilde devam etmektedir ⁽¹⁾. O zamandan beri, bağışıklık sistemi ile tümör büyümesi ve gelişimi arasındaki kesişimin artması, tüm kanser türlerinde geniş terapötik ilerlemelere yol açmıştır.

Kanser hastalarının tedavisinde son zamanlarda çeşitli immünoterapötiklerin uygulanması, devrim niteliğinde değişimlere neden olmuştur. Bununla birlikte, yanıt oranları hala yaklaşık % 20 ila %40 arasında sınırlı olup, kemoterapi, radyoterapi, epigenetik modülatörler ve hedefe yönelik tedaviler gibi geleneksel tedaviler ile immünoterapi kombinasyonlarının, anti-tümör bağışıklığını arttırmak için bir seçenek olabileceğini düşündürmektedir. İmmünoterapilerle kombinasyon halinde farklı immün olmayan bazı tedavilerin, bağışıklık baskılayıcı tümör mikro-ortamını yeniden programlayabileceği ve tümör hücrelerinin immünojenikliğini artırabileceği ve bunun da daha iyi terapötik etkinliğe ve hasta sonuçlarına yol açabileceği düşünülmektedir. Bu farklı terapilerin immünoterapilere eklenmesiyle elde edilen artan objektif cevapların çeşitli örnekleri olmasına rağmen, hastalar için klinik faydayı en üst düzeye çıkarmak amacıyla kontrol noktası inhibitör kombinasyonlarının rasyonel ve kanıta dayalı tedavi stratejilerine dönük çalışmalar yoğunluk kazanmıştır. Özellikle bağışıklık kontrol noktası (immune checkpoint:iCP) moleküllerine karşı oluşturulan T lenfosit antijen 4 (CTLA-4)

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infiltrasyonu ve PD-L1 gibi iCP'lerin ekspresyonu, kontrol noktası blokajı cevabı için öngörücü olduğu tartışılmıştır. Buna ek olarak, iCPI'nin diğer terapilerle kombinasyonlarının listesi geniştir ve iCPI kombinasyon terapilerinin potansiye-line rağmen, daha fazla hasta için hayatta kalma yararı sağlamak için immünoterapi kombinasyonları uygulanan hastaları seçmek için biyobelirteçleri tanımlama ve azaltılmış yan etki ihtiyacının altını çizmek gerekmektedir. Son çalışmalarda, TMB, iCPI ekspresyonu ve immün hücre infiltrasyonu, konak genetik, mikrosatellit instabilitesi, neo-antijen kaybının, TME'nin hücresel olmayan bileşimi ve mikrobiyom analizi önerilmektedir. Bu nedenle, standartlaştırılmış ve optimize edilmiş (bağışıklık) izlemenin yanısıra bağışıklık tepkisinin dinamiklerini, posttranslasyonel modifikasyonları, bağışıklık ve tümör hücrelerinin düzeni , ayrıca TME'nin, hipoksi ve pH değeri gibi fiziksel faktörler immünoterapileri uyarlamak için çok önemlidir. Bu zorlukların üstesinden gelmek ve yeni ajanların uygulanması ve kombinasyon stratejiler şu anda iCPI tedavisinde etkinliklerini artırmak ve dirençleri önlemek için ana araştırma odağıdır.

Kemoterapinin immünostimülatör etkileri hakkında çok fazla araştırma yapılmış olsada, dozaj ve program, uzun süreli bağışıklık belleği ve kemoterapi ve immünoterapi kombinasyonları söz konusu olduğunda henüz çok az şey öğrenilmiştir. Yeni kombinasyon tedavilerinin geliştirilmesi, immünoterapi uygulamalarını genişletecek ve daha erişilebilir hale getirecektir.

KAYNAKLAR

1. Coley WB The treatment of malignant tumors by repeated inoculations of erysipelas: with a report of ten original cases Am J Med Sci. 1893;105:487.
2. Mellman I, Coukos G, Dranoff G. Cancer immunotherapy comes of age. Nature. (2011) 480:480–9. doi: 10.1038/nature10673
3. Migden MR, Rischin D, Schmults CD, Guminski A, Hauschild A, Lewis KD, et al. PD-1 blockade with cemiplimab in advanced cutaneous squamous-cell carcinoma. N Engl J Med. (2018) 379:341–51. doi: 10.1056/NEJMoa1805131
4. Cemiplimab Approved for Treatment of CSCC (2018). CancerDiscov. 8:OF2.doi: 10.1158/2159-8290.CD-NB2018-140
5. Cogdill AP, Andrews MC, Wargo JA. Hallmarks of response to immune checkpoint blockade. Br J Cancer. (2017) 117:1–7. doi: 10.1038/bjc.2017.136
6. MacGregor HL, Ohashi PS. Molecular pathways: evaluating the potential for B7-H4 as an immunoregulatory target. Clin Cancer Res. (2017) 23:2934–41. doi: 10.1158/1078-0432.CCR-15-2440
7. Yun S, Vincelette ND, Green MR, Wahner Hendrickson AE, Abraham I. Targeting immune checkpoints in unresectable metastatic cutaneous melanoma: a systematic review and meta-analysis of anti-CTLA-4 and anti- PD-1 agents trials. Cancer Med. (2016) 5:1481–91. doi: 10.1002/cam4.732
8. Lu J, Lee-Gabel L, Nadeau MC, Ferencz TM, Soefje SA. Clinical evaluation of compounds targeting PD-1/PD-L1 pathway for cancer immunotherapy. J Oncol Pharm Pract. (2015) 21:451–67. doi: 10.1177/1078155214538087

9. Baumeister SH, Freeman GJ, Dranoff G, Sharpe AH. Coinhibitory pathways in immunotherapy for cancer. *Annu Rev Immunol.* (2016) 34:539–73. doi: 10.1146/annurev-immunol-032414-112049
10. Melero I, Berman DM, Aznar MA, Korman AJ, Perez Gracia JL, Haanen J. Evolving synergistic combinations of targeted immunotherapies to combat cancer. *Nat Rev Cancer.* (2015) 15:457–72. doi: 10.1038/nrc3973
11. Mahoney KM, Rennert PD, Freeman GJ. Combination cancer immunotherapy and new immunomodulatory targets. *Nat Rev Drug Discov.* (2015) 14:561–84. doi: 10.1038/nrd4591
12. Smyth MJ, Ngiew SF, Ribas A, Teng MW. Combination cancer immunotherapies tailored to the tumour microenvironment. *Nat Rev Clin Oncol.* (2016) 13:143–58. doi: 10.1038/nrclinonc.2015.209
13. Morrissey KM, Yuraszeck TM, Li CC, Zhang Y, Kasichayanula S. Immunotherapy and novel combinations in oncology: current landscape, challenges, and opportunities. *Clin Transl Sci.* (2016) 9:89–104. doi: 10.1111/cts.12391
14. Bates SE. Refining immunotherapy approvals. *Clin Cancer Res.* (2017) 23:4948–9. doi: 10.1158/1078-0432.CCR-17-2025
15. Lhuillier C, Vanpouille-Box C, Galluzzi L, Formenti SC, Demaria S. Emerging biomarkers for the combination of radiotherapy and immune checkpoint blockers. *Semin Cancer Biol.* (2018) 52(Pt. 2):125–34. doi: 10.1016/j.semcancer.2017.12.007
16. Lesterhuis WJ, Bosco A, Millward MJ, Small M, Nowak AK, Lake RA. Dynamic versus static biomarkers in cancer immune checkpoint blockade: unravelling complexity. *Nat Rev Drug Discov.* (2017) 16:264–72. doi: 10.1038/nrd.2016.233
17. Pulluri B, Kumar A, Shaheen M, Jeter J, Sundararajan S. Tumor microenvironment changes leading to resistance of immune checkpoint inhibitors in metastatic melanoma and strategies to overcome resistance. *Pharmacol Res.* (2017) 123:95–102. doi: 10.1016/j.phrs.2017.07.006
18. J.A. Joyce, D.T. Fearon, T cell exclusion, immune privilege, and the tumor microenvironment, *Science* 348 (6230) (2015) 74–80.
19. D.S. Chen, I. Mellman, Oncology meets immunology: the cancer-immunity cycle, *Immunity* 39 (1) (2013) 1–10.
20. T. Karasaki, et al., An immunogram for the cancer-immunity cycle: towards personalized immunotherapy of lung cancer, *J. Thorac. Oncol.* 12 (5) (2017) 791–803.
21. T. Weiss, M. Weller, P. Roth, Immunological effects of chemotherapy and radiotherapy against brain tumors, *Expert Rev. Anticancer Ther.* 16 (10) (2016) 1087–1094.
22. J. Qiao, Z. Liu, Y.X. Fu, Adapting conventional cancer treatment for immunotherapy, *J. Mol. Med. (Berl.)* 94 (5) (2016) 489–495.
23. Brahmer J, Reckamp KL, Baas P, et al. Nivolumab versus docetaxel in advanced squamous-cell non-small-cell lung cancer. *N Engl J Med* 2015;373:123-35. 10.1056/NEJMoa1504627. 26028407
24. Michael A. Postow, M.D. Immune-Related Adverse Events Associated with Immune Checkpoint Blockade *N Engl J Med* 2018;378:158-68. DOI: 10.1056/NEJMra1703481
25. Gandhi L, Rodríguez-Abreu D, Gadgeel S, et al. KEYNOTE-189 Investigators. Pembrolizumab plus chemotherapy in metastatic non-small-cell lung cancer. *N Engl J Med* 2018;378:2078-92. 10.1056/NEJMoa1801005. 29658856
26. D.L. Barber, E.J. Wherry, D. Masopust, B. Zhu, J.P. Allison, A.H. Sharpe, G.J. Freeman, R. Ahmed, Restoring function in exhausted CD8 T cells during chronic viral infection, *Nature* 439 (2006) 682–687, <https://doi.org/10.1038/nature04444>.
27. M.M. Huerter, A.K. Ganti, Predictive biomarkers for immune checkpoint inhibitor therapy: we need to keep searching, *J. Thorac. Dis.* 10 (2018) S2195–S2197, <https://doi.org/10.21037/jtd.2018.06.144>.
28. K.G. Anderson, I.M. Stromnes, P.D. Greenberg, Obstacles posed by the tumor microenvironment to T cell activity: a case for synergistic therapies, *Cancer Cell* 31 (2017) 311–325, <https://doi.org/10.1016/j.ccell.2017.02.008>.Obstacles.

29. R.W. Jenkins, D.A. Barbie, K.T. Flaherty, Mechanisms of resistance to immune ncheckpoint inhibitors, *Br. J. Cancer* 118 (2018) 9–16, <https://doi.org/10.1038/bjc.2017.434>.
30. L. Bracci, G. Schiavoni, A. Sistigu, F. Belardelli, Immune-based mechanisms of cytotoxic chemotherapy: implications for the design of novel and rationale-based combined treatments against cancer, *Cell Death Differ.* 21 (2014) 15–25, <https://doi.org/10.1038/cdd.2013.67>.
31. T. Sugimura, The principles of cancer treatment—changes in chemotherapy, *Gan To Kagaku Ryoho* 29 (7) (2002) 1263–1278.
32. O. Kepp, et al., Molecular determinants of immunogenic cell death elicited by anticancer chemotherapy, *Cancer Metastasis Rev.* 30 (1) (2011) 61–69.
33. M. Obeid, et al., Ecto-calreticulin in immunogenic chemotherapy, *Immunol. Rev.* 220 (2007) 22–34.
34. G. Chen, L.A. Emens, Chemoimmunotherapy: reengineering tumor immunity, *Cancer Immunol. Immunother.* 62 (2) (2013) 203–216.
35. A.M. McDonnell, et al., Tumor-infiltrating dendritic cells exhibit defective crosspresentation of tumor antigens, but is reversed by chemotherapy, *Eur. J. Immunol.* 45 (1) (2015) 49–59.
36. T. Browder, et al., Antiangiogenic scheduling of chemotherapy improves efficacy against experimental drug-resistant cancer, *Cancer Res.* 60 (7) (2000) 1878–1886.
37. F. Bertolini, et al., Maximum tolerable dose and low-dose metronomic chemotherapy have opposite effects on the mobilization and viability of circulating endothelial progenitor cells, *Cancer Res.* 63 (15) (2003) 4342–4346.
38. G. Klement, et al., Continuous low-dose therapy with vinblastine and VEGF receptor- 2 antibody induces sustained tumor regression without overt toxicity, *J. Clin. Investig.* 105 (8) (2000) R15–R24.
39. Y.K. Chae, A. Arya, W. Iams, M. Cruz, S. Chandra, J. Choi, F. Giles, Current landscape and future of dual anti- CTLA4 and PD-1/PD-L1 blockade immunotherapy in cancer; lessons learned from clinical trials with melanoma and non- small cell lung cancer (NSCLC), *J. Ultrasound Med.* 13 (1994) 807–808, <https://doi.org/10.7863/jum.1994.13.10.807>.
40. Y.L. Chen, M.C. Chang, W.F. Cheng, Metronomic chemotherapy and immunotherapy in cancer treatment, *Cancer Lett.* 400 (2017) 282–292.
41. I. Angulo, et al., Nitric oxide-producing CD11b(+)Ly-6G(Gr-1)(+)CD31(ER-MP12) (+) cells in the spleen of cyclophosphamide-treated mice: implications for T-cell responses in immunosuppressed mice, *Blood* 95 (1) (2000) 212–220.
42. M. Mkrtychyan, Y.G. Najjar, E.C. Raulfs, M.Y. Abdalla, R. Samara, R. Rotem- Yehudar, L. Cook, S.N. Khleif, Anti-PD-1 synergizes with cyclophosphamide to induce potent anti-tumor vaccine effects through novel mechanisms, *Eur. J. Immunol.* 41 (2011) 2977–2986,
43. I.C. Lien, S.M. Pollack, Limited activity of metronomic cyclophosphamide and pembrolizumab for soft tissue sarcomas, *Transl. Gastroenterol. Hepatol.* 2018 (2018) 3–5,
44. R. De Souza, et al., Chemotherapy dosing schedule influences drug resistance development in ovarian cancer, *Mol. Canc. Therapeut.* 10 (7) (2011) 1289–1299.
45. X. Qin, C. Liu, Y. Zhou, G. Wang, Cisplatin induces programmed death-1-ligand 1(PD-L1) over-expression in hepatoma H22 cells via Erk/MAPK signaling pathway, *Cell. Mol. Biol. (Noisy-Le-Grand)* 56 (Suppl) (2010) OL1366–72
46. L. Tran, C.T. Allen, R. Xiao, E. Moore, R. Davis, S.-J. Park, K. Spielbauer, C. Van Waes, N.C. Schmitt, Cisplatin alters antitumor immunity and synergizes with PD- 1/PD-L1 inhibition in head and neck squamous cell carcinoma, *Cancer Immunol. Res.* 5 (2017) 1141–1151.
47. H. Wimberly, J.R. Brown, K. Schalper, H. Haack, M.R. Silver, C. Nixon, V. Bossuyt, L. Pusztai, D.R. Lannin, D.L. Rimm, PD-L1 expression correlates with tumor-infiltrating lymphocytes and response to neoadjuvant chemotherapy in breast cancer, *Cancer Immunol. Res.* 3 (2015) 326–332, <https://doi.org/10.1158/2326-6066.CIR-14-0133>.
48. H. El-Saghire, et al., Low doses of ionizing radiation induce immune-stimulatory responses in isolated human primary monocytes, *Int. J. Mol. Med.* 32 (6) (2013) 1407–1414.

49. A.D. Garg, et al., Immunogenic cell death, DAMPs and anticancer therapeutics: an emerging amalgamation, *Biochim. Biophys. Acta* 1805 (1) (2010) 53–71.
50. P.M. Gallo, S. Gallucci, The dendritic cell response to classic, emerging, and homeostatic danger signals. Implications for autoimmunity, *Front. Immunol.* 4 (2013) 138
51. Marleen Kok, Hugo M. Horlings Adaptive phase II randomized trial of nivolumab after induction treatment in triple negative breast cancer (TONIC trial): Final response data stage I and first translational data. *ClinicalTrials.gov Identifier: NCT02499367*
52. S. Desai, et al., Cytokine profile of conditioned medium from human tumor cell lines after acute and fractionated doses of gamma radiation and its effect on survival of bystander tumor cells, *Cytokine* 61 (1) (2013) 54–62.
53. S. Matsumura, et al., Radiation-induced CXCL16 release by breast cancer cells attracts effector T cells, *J. Immunol.* 181 (5) (2008) 3099–3107.
54. L.A. Emens, et al., Long-term clinical outcomes and biomarker analyses of atezolizumab therapy for patients with metastatic triple-negative breast cancer: a phase 1 study, *JAMA Oncol* 5 (1) (2018) 74–82.
55. S. Adams, et al., Atezolizumab plus nab-paclitaxel in the treatment of metastatic triple-negative breast cancer with 2-year survival follow-up: a phase 1b clinical trial, *JAMA Oncol* 5 (3) (2018) 334–342.
56. P. Schmid, et al., Atezolizumab and nab-paclitaxel in advanced triple-negative breast cancer, *N. Engl. J. Med.* 379 (22) (2018) 2108–2121.
57. L. Horn, et al., First-line atezolizumab plus chemotherapy in extensive-stage smallcell lung cancer, *N. Engl. J. Med.* 379 (23) (2018) 2220–2229.
58. Wei-Di Yu, Guan Sun , Mechanisms and therapeutic potentials of cancer immunotherapy in combination with radiotherapy and/or chemotherapy, Received 2 January 2019; Received in revised form 8 February 2019; Accepted 13 February 2019
59. C. Ratti, L. Botti, V. Cancila, S. Galvan, I. Torselli, C. Garofalo, M.C. Manara, L. Bongiovanni, C.F. Valenti, A. Burocchi, M. Parenza, B. Cappetti, S. Sangaletti, C. Tripodo, K. Scotlandi, M.P. Colombo, C. Chiodoni, Trabectedin overrides osteosarcoma differentiative block and reprograms the tumor immune environment enabling effective combination with immune checkpoint inhibitors, *Clin. Cancer Res.* 23 (2017) 5149–5161,
60. L. L., X. X., B. Z., R. Z., H. J., X. W., Combined PD-1 blockade and GITR triggering induce a potent antitumor immunity in murine cancer models and synergizes with chemotherapeutic drugs, *J. Transl. Med.* 12 (2014) 1–11
61. W. Wang, L. Wu, J. Zhang, H. Wu, E. Han, Q. Guo, Chemoimmunotherapy by combining oxaliplatin with immune checkpoint blockades reduced tumor burden in colorectal cancer animal model, *Biochem. Biophys. Res. Commun.* 487 (2017) 1–7,
62. M. Jure-Kunkel, G. Masters, E. Girit, G. Dito, F. Lee, J.T. Hunt, R. Humphrey, Synergy between chemotherapeutic agents and CTLA-4 blockade in preclinical tumor models, *Cancer Immunol. Immunother.* 62 (2013) 1533–1545,