

BÖLÜM 6

KÜLTÜR BALIKLARINDA İMMÜNOSTİMÜLAN KULLANIMI

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GİRİŞ

Su ürünleri üretim sektörü, özellikle balık yetiştiriciliği alanı, besin ihtiyacını karşılamak için dünya genelinde nüfus artışına paralel olarak gelişmektedir. Dünya nüfusunun 2050 yılına kadar yaklaşık 10 milyar olması beklenmektedir. Su ürünleri üretim sektörünün de büyümesini sürdüreceği ve kilit rol üstlenmeye devam edeceği öngörülmektedir (1). Küresel su ürünleri yetiştiriciliği son 40 yıldır istikrarlı bir şekilde artmaktadır ve 2022 yılında 94.415.401 ton ile avcılık ile elde edilen toplam miktarı (92.320.427 ton) geride bırakmıştır (2). TÜİK verilerine göre 2023 yılında Türkiye’de 553.862 ton su ürünleri yetiştiriciliği yapılmıştır (3). Veriler incelendiğinde su ürünleri yetiştiriciliğinin dünyayla paralel olarak ülkemizde de özellikle 2011 yılından sonra sürekli artan ve büyüyen bir sektör olarak karşımıza çıktığı görülmektedir. Sektörde stoklama ve yem üretimindeki teknolojik gelişmeler sayesinde yoğun yetiştiricilik sınırları zorlanmakta ve yoğun stoklama ile yetiştiricilik alanından en yüksek verim alınmaya çalışılmaktadır. Yüksek stok yoğunluğu, alternatif yem kaynaklarından yüksek yararlanım ve yetiştiricilik yapılan sucul ortamın kalitesinin korunması yetiştiricilik sistemlerinde yaygın hayvancılık uygulamaları haline gelmiştir (4). Bu durum yetiştiricilik açısından başarılı sonuçlar alınmasını sağlamış olsa da hastalık, stres kaynaklı sağlık riskleri ve ortam kalitesinin düşmesi gibi problemleri de beraberinde getirmiştir.

Stres etmenleri bağışıklık sistemini olumsuz önde etkilemekte ve canlıyı tehditlere karşı daha zayıf düşürmektedir. Hastalık kaynaklı ölümlerin balık yetiştiriciliğinde %50 gibi büyük bir oranda üretim kaybına sebep olduğu bildirilmiştir (5,

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şıklık sisteminin zayıflamasına ve büyüme performansının düşmesine sebep olduğu, yem çevirim oranlarında gerilemeye sebep olduğu bildirilmiştir (89).

SONUÇ

Sürekli artış gösteren insan nüfusuna paralel olarak nitelikli besin ihtiyacına olan talep de artış göstermektedir. Bu gereksinimi önemli oranla karşılayan su ürünleri yetiştiricilik sektörünün hedefi büyümeye devam ederek mevcut alanlardaki üretimi miktarını arttırmak, daha sürdürülebilir yemler kullanmak, hastalıklardan korunmak ve sağlıklı bireyler yetiştirmektir.

Su ürünleri kültüründe karasal yetiştiricilikten farklı olarak bir olumsuzluk durumunda canlılara müdahale kolay değildir, yoğun iş gücü ve operasyonel hareket gerektirir. Balıkların aynı su ortamında etkileşiminden dolayı bir hastalık belirtisi ortaya çıktığında kuluçka süresince bulaş çoğunlukla gerçekleşmiş ve hastalık yayılmıştır. Bu durum kitlesel balık kayıplarıyla gıda arzını etkilemekte ve işletmeler için ağır ekonomik kayıplar anlamına gelmektedir.

Bu durumlardan dolayı immünostimülanlar bir destek olarak araştırmacı ve yetiştiricilerin ilgisini çekmiştir. Bağışıklık sisteminin uyarılması ve desteklenmesi ile canlı kayıplarında azalma, hastalık direnci ve yem alımında iyileşme gibi ürün kaybına engel olacak gelişmeler immünostimülan grubunda yer alan farklı kimyasal ya da mikrobiyolojik ajanlarla sağlanmıştır (9, 48, 49).

İmmünostimülanlar günümüz yoğun yetiştiricilik koşullarında önemli birer destek olarak görünmekte olup, yetiştiricilik hedeflerinin artış planlanması doğrultusunda zamanla daha önemli hale geleceği düşünülmektedir. Yine çalışmalar göstermiştir ki bir patojenin ya da canlı sağlığını olumsuz etkileyen herhangi bir koşulun olumsuz etkilerini sadece immünostimülanlarla ortadan kaldırmak mümkün değildir. Ayrıca, immünostimülan etki gösteren moleküllerin ya da bileşiklerin tür ve yaşam evresi ile dozlarının da değişim gösterdiği unutulmamalıdır. Ancak üretimde artış ve sürdürülebilirlik açısından özellikle yoğun yetiştiricilik koşullarında alternatif değil, diyet formüllerinin bir parçası olarak düşünülmesi üretimin lehine görünmektedir.

KAYNAKLAR

1. Santhosh AS, Umesh MA. Strategic review on use of polyhydroxyalkanoates as an immunostimulant in aquaculture. *Applied Food Biotechnology*; 2021;8(1), 1-18.
2. Food and Agriculture Organization of the United Nations, 2024. "In Brief to The State of World Fisheries and Aquaculture 2024", Blue Transformation in action. Rome, 40 syf. <https://doi.org/10.4060/cd0690en>
3. TÜİK. Su ürünleri İstatistikleri. (08.10.2025 tarihinde <https://data.tuik.gov.tr/Bulten/Index?p=-Fishery-Products-2024-54193> adresinden ulaşılmıştır).

4. Mastan SA. Use of immunostimulants in aquaculture disease management. *International Journal of Fisheries and Aquatic Studies*; 2015; 2(4), 277-280.
5. Gabriel NN. Review on the progress in the role of herbal extracts in tilapia culture. *Cogent Food Agri.*, 2019;5: 1619651.
6. Gupta N, Rani Kar S, Chakraborty A. A review on medicinal plants and immune status of fish. *Egyptian Journal of Aquatic Biology and Fisheries*; 2021;25(2), 897-912.
7. Ergönül M.B, Yavuzcan H, Altindag A. Balık Sağlığı ve Immunostimulanların Kullanımı/Fish Health and the Use of Immunostimulants. *Journal of Fisheries Sciences*; 2012; 6(3), 188.
8. Vijayaram S, Ringø E, Zuurro A. et al. Beneficial roles of nutrients as immunostimulants in aquaculture: A review. *Aquaculture and Fisheries*; 2024; 9(5), 707-720.
9. Bairwa MK, Jakhar JK, Satyanarayana Y. Et al. Animal and plant originated immunostimulants used in aquaculture. *Journal of Natural Product and PlantResources*; 2012;2(3), 397-400.
10. Vallejos-Vidal, E., Reyes-López, F., Teles, M., & MacKenzie, S. (2016). The response of fish to immunostimulant diets. *Fish & Shellfish Immunology*, 56, 34-69.
11. Ganguly S, Paul I, Mukhopadhyay SK. Application and effectiveness of immunostimulants, probiotics, and prebiotics in aquaculture: a review. *The Israeli Journal of Aquaculture – Bami-dgeh*, 2010;62(3), 130-138.
12. Magnadottir B. Innate immunity of fish (overview). *Fish & Shellfish Immunology*; 2006; 20(2), 137–151. DOI:10.1016/j.fsi.2004.09.006
13. Reyes-Cerpa S, Maisey K, Reyes-Lepez F, et al. Fish Cytokines and Immune Response. In *New Advances and Contributions to Fish Biology 2012*; (pp. 3–57). InTech. DOI:10.5772/53504.
14. Shahbazi S, Bolhassani A. Immunostimulants: types and functions. *Journal of Medical Microbiology and Infectious Diseases*; 2016;4(3), 45-51.
15. Wang W, Sun J, Liu C, Xue Z. Application of immunostimulants in aquaculture: current knowledge and future perspectives. *Aquaculture Research*; 2017;48(1), 1-23.
16. Sakai M. Current research status of fish immunostimulants. *Aquaculture*; 1999;172: 63-92.
17. Ringø E, Olsen RE, Vecino JG, et al. Use of immunostimulants and nucleotides in aquaculture: a review. *J Mar Sci Res Dev.*; 2012;2(1), 104.
18. Bricknell I, Dalmo RA. The use of immunostimulants in fish larval aquaculture. *Fish Shellfish Immunology*; 2005;19: 457-472.
19. Volman JJ, Ramakers JD, Plat J. Dietary modulation of immune function by β -glucans. *Physiol-Behavior.*; 2008;94: 276-284.
20. Cook MT, Hayball PJ, Hutchinson W. Et al. Administration of a commercial immunostimulant preparation, EcoActiva™ as a feed supplement enhances macrophage respiratory burst and the growth rate of snapper (*Pagrus auratus*, Sparidae (Bloch and Schneider) in winter. *Fish & Shellfish Immunology*; 2003;14(4), 333-345.
21. Jeney G, Anderson DP. Glucan injection or bath exposure given alone or in combination with a bacterin enhance the non-specific defence mechanisms in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*; 1993;116, 315–329.
22. Reverter M, Bontemps N, Lecchini D, et al. Use of plant extracts in fish aquaculture as an alternative to chemotherapy: current status and future perspectives. *Aquac.*, 2014;433: 50- 61.
23. Van Hai N. The use of medicinal plants as immunostimulants in aquaculture: A review. *Aquac.*; 2015;446: 88-96.
24. Ghosh K, Ray AK, Ringø E. Applications of plant ingredients for tropical and subtropical freshwater finfish: possibilities and challenges. *Rev. Aquac.*; 2018;11: 793-815
25. Jian J, Wu Z. Influences of traditional Chinese medicine on non-specific immunity of Jian carp (*Cyprinus carpio* var. Jian). *Fish. Shellfish. Immunol.*; 2004;16: 185-191.
26. Ashokkumar K, Selvaraj K, Muthukrishnan SD. *Cynodon dactylon* (L.) Pers.: An updated review of its phytochemistry and pharmacology. *J Med Plants Res.*; 2013;7(48), 3477-3483.
27. Habotta OA, Dawood MA, Kari ZA et al. Antioxidative and immunostimulant potential of fruit derived biomolecules in aquaculture. *Fish & shellfishimmunology*; 2022;130, 317-322.
28. Harikrishnan R, Thamizharasan S, Devi G, et al. Dried lemon peel enriched diet improves anti-

- oxidant activity, immune response and modulates immuno-antioxidant genes in *Labeo rohita* against *Aeromonas sorbia*. *Fish & Shellfish Immunology*; 2020;106, 675-684.
29. Mousavi S, Sheikhzadeh N, Hamidian G, et al. Changes in rainbow trout (*Oncorhynchus mykiss*) growth and mucosal immune parameters after dietary administration of grape (*Vitis vinifera*) seed extract. *Fish Physiology and Biochemistry*; 2021;47, 547-563.
 30. Beltran JMG, Espinosa C, Guardiola FA., et al. Dietary dehydrated lemon peel improves the immune but not the antioxidant status of gilthead seabream (*Sparus aurata* L.). *Fish & shellfish immunology*; 2017;64, 426-436.
 31. Kilercioğlu S. Balıklarda bağışıklık sistemi, mukozal bağışıklık ve IL-1 β , IL-18 ve TNF- α pro-enflamatuar sitokinlerinin işlevleri. *Ege Journal of Fisheries and Aquatic Sciences*; 2021;38(1), 125-134.
 32. Lee BJ, Kim SS, Song JW, et al. Effects of dietary supplementation of citrus by-products fermented with a probiotic microbe on growth performance, innate immunity and disease resistance against *Edwardsiella tarda* in juvenile olive flounder, *Paralichthys olivaceus* (Temminck & Schlegel). *Journal of Fish Diseases*, 2013;36(7), 617-628.
 33. Giri SS, Jun JW, Sukumaran V, Park SC. Dietary administration of banana (*Musa acuminata*) peel flour affects the growth, antioxidant status, cytokine responses, and disease susceptibility of rohu, *Labeo rohita*. *Journal of immunology research*, 2016(1), 4086591.
 34. Harikrishnan R, Devi G, Van Doan H et al. Impact of grape pomace flour (GPF) on immunity and immune-antioxidant-anti-inflammatory genes expression in *Labeo rohita* against *Flavobacterium columnaris*. *Fish & Shellfish Immunology*; 2021;111, 69-82.
 35. Mohan K, Rajan DK, Ganesan AR et al. Chitin, chitosan and chitooligosaccharides as potential growth promoters and immunostimulants in aquaculture: A comprehensive review. *International Journal of Biological Macromolecules*, 2023;126285.
 36. Gopalakannan A, Arul V. Immunomodulatory effects of dietary intake of chitin, chitosan and levamisole on the immune system of *Cyprinus carpio* and control of *Aeromonas hydrophila* infection in ponds. *Aquaculture*; 2006;255, 179-187.
 37. Wang SH, Chen JC. The protective effect of chitin and chitosan against *Vibrio alginolyticus* in white shrimp *Litopenaeus vannamei*. *Fish and Shellfish Immunology*; 2005;19, 191-204.
 38. Srirengaraj V, Razafindralambo HL, Rabetafika HN, et al. Synbiotic Agents and Their Active Components for Sustainable Aquaculture: Concepts, Action Mechanisms, and Applications. *Biology*; 2023;12(12), 1498.
 39. Li X, Ringø E, Hoseinifar SH et al. The adherence and colonization of microorganisms in fish gastrointestinal tract. *Reviews in Aquaculture*; 2019;11(3), 603-618.
 40. Cavalcante RB, Telli GS, Tachibana L, et al. Probiotics, Prebiotics and Synbiotics for Nile tilapia: Growth performance and protection against *Aeromonas hydrophila* infection. *Aquaculture Reports*; 2020;17, 100343.
 41. Devi G, Harikrishnan R, Paray BA, et al. Effect of symbiotic supplemented diet on innate-adaptive immune response, cytokine gene regulation and antioxidant property in *Labeo rohita* against *Aeromonas hydrophila*. *Fish & shellfish immunology*; 2019;89, 687-700.
 42. Socol CR, Vandenberghe LDS, Spier MR, et al. The potential of probiotics: a review. *Food Technol. Biotechnol.*, 2010;48(4), 413-434.
 43. Rohani ME, Islam SM, Hossain MK, et al. Probiotics, prebiotics and synbiotics improved the functionality of aquafeed: Upgrading growth, reproduction, immunity and disease resistance in fish. *Fish & shellfish immunology*, 2022;120, 569-589.
 44. Liu H, Li Z, Tan B, et al. Isolation of a putative probiotic strain S12 and its effect on growth performance, non-specific immunity and disease-resistance of white shrimp, *Litopenaeus vannamei*. *Fish & shellfish immunology*, 2014;41(2), 300-307.
 45. Mamun MAA, Nasren S, Rathore SS, et al. Assessment of probiotic in aquaculture: functional changes and impact on fish gut. *Microbiology Research Journal International*; 2019;29(1), 1-10.
 46. Merrifield DL, Dimitroliou A, Foey A, et al. The current status and future focus of probiotic and prebiotic applications for salmonids. *Aquaculture*, 2010;302:1-18.

47. Fuller R. Probiotics in man and animals. The Journal of applied bacteriology, 1989;66(5), 365-378.
48. Jahan N, Islam SM, Rohani MF, et al. Probiotic yeast enhances growth performance of rohu (*Labeo rohita*) through upgrading hematology, and intestinal microbiota and morphology. Aquaculture, 2021;545, 737243.
49. Haque MM, Hasan NA, Eltholth MM, et al. Assessing the impacts of in-feed probiotic on the growth performance and health condition of pangasius (*Pangasianodon hypophthalmus*) in a farm trial. Aquaculture reports, 2021;20, 100699.
50. Kilercioglu S, Koşker AR, Evliyaoglu E. Public health risk assessments associated with heavy metal levels in panga fish fillets imported from Vietnam. International Journal of Agriculture Environment and Food Sciences, 2022;6(4), 568-578.
51. Srednicka P, Juszczuk-Kubiak E, Wójcicki M, Akimowicz M. et al. Probiotics as a biological detoxification tool of food chemical contamination: A review. Food and Chemical Toxicology, 2021;153, 112306.
52. Akhter N, Wu B, Memon AM. Et al. Probiotics and prebiotics associated with aquaculture: a review. Fish & shellfish immunology, 2015;45(2), 733-741.
53. Newaj-Fyzul A, Al-Harbi AH, Austin, B. Developments in the use of probiotics for disease control in aquaculture. Aquaculture; 2014; 431, 1-11.
54. Manning TS, Gibson GR. Prebiotics. Best practice & research clinical gastroenterology, 2004;18(2), 287-298.
55. Hoseinifar SH, Mirvaghefi A, Amoozegar MA., et al. Modulation of innate immune response, mucosal parameters and disease resistance in rainbow trout (*Oncorhynchus mykiss*) upon syn-biotic feeding. Fish Shellfish Immunol., 2015;45: 27–32.
56. Hasan T, Jang WJ, Tak JY, Lee BJ, et al. Effects of *Lactococcus lactis* subsp. *lactis* I2 with b-glucan oligosaccharides on growth, innate immunity and streptococcosis resistance in olive flounder (*Paralichthys olivaceus*). J. Microbiol. Biotechnol.; 2018;28:1433–1442
57. Ridwanudin A, Anggorowati DA, Indriana LF, Fahmi V. Current status of immunostimulant agents in tropical aquaculture: a review. Aquaculture, Aquarium, Conservation & Legislation, 2022;15(4), 1908-1925.
58. Low C, Wadsworth S, Burrells C, Secombes CJ. Expression of immune genes in turbot (*Scophthalmus maximus*) fed a nucleotide-supplemented diet. Aquaculture, 2003;221(1-4), 23-40.
59. Burrells C, Williams PD, Southgate PJ, Wadsworth, S.L., Dietary nucleotides: a novel supplement in fish feeds: 2. Effects on vaccination, salt water transfer, growth rates and physiology of atlantic salmon (*Salmo salar* L.). Aquaculture, 2001;199, 171 – 184.
60. Selim KM, Reda RM, Mahmoud R, El-Araby, I. E. Effects of nucleotides supplemented diets on growth performance and expressions of ghrelin and insulin-like growth factor genes in Nile tilapia, *Oreochromis niloticus*. Journal of Applied Aquaculture, 2020;32(2), 157-174.
61. Li P, Gatlin III DM. Nucleotide nutrition in fish: current knowledge and future applications. Aquaculture, 2006;251(2-4), 141-152.
62. Dawood MA, Koshio S, Esteban MÁ. Beneficial roles of feed additives as immunostimulants in aquaculture: a review. Reviews in Aquaculture, 2018;10(4), 950-974.
63. Hossain MS, Koshio S, Kestemont P. Recent advances of nucleotide nutrition research in aquaculture: a review. Reviews in Aquaculture, 2020;12(2), 1028-1053.
64. Craig SR, Helfrich LA, Kuhn D, Schwarz MH. Understanding fish nutrition, feeds, and feeding. 2017. <https://vtechworks.lib.vt.edu/server/api/core/bitstreams/24c04f50-8d2f-4b2d-9f8a-9e-c3684537a1/content>
65. Buentello JA, Gatlin DM. Effects of elevated dietary arginine on resistance of channel catfish to exposure to *Edwardsiella ictaluri*. Journal of Aquatic Animal Health; 2001;13:194–201.
66. Hoseini SM, Ahmad Khan M, Yousefi M, et al. Roles of arginine in fish nutrition and health: insights for future researches. Reviews in Aquaculture, 2020;12(4), 2091-2108.
67. Natnan ME, Low CF, Chong CM. et al. Oleic acid as potential immunostimulant in metabolism pathways of hybrid grouper fingerlings (*Epinephelus fuscoguttatus* × *Epinephelus lanceolatus*)

- infected with *Vibrio vulnificus*. Scientific Reports, 2023;13(1), 12830.
68. Machado M, Azeredo R, Diaz-Rosales P, et al. Dietary tryptophan and methionine as modulators of European seabass (*Dicentrarchus labrax*) immune status and inflammatory response. Fish & shellfish immunology, 2015;42(2), 353-362.
69. Tejpal CS, Pal AK, Sahu NP, et al. Dietary supplementation of L-tryptophan mitigates crowding stress and augments the growth in *Cirrhinus mrigala* fingerlings. Aquaculture; 2009;293(3-4), 272-277.
70. Azeredo R, Machado M, Guardiola FA, et al. Local immune response of two mucosal surfaces of the European seabass, *Dicentrarchus labrax*, fed tryptophan- or methionine-supplemented diets. Fish Shellfish Immunology; 2017;70, 76-86. DOI:10.1016/j.fsi.2017.09.016.
71. Machado M, Moura J, Peixoto D, et al. Dietary methionine as a strategy to improve innate immunity in rainbow trout (*Oncorhynchus mykiss*) juveniles. General and Comparative Endocrinology, 2021;302, 113690.
72. Zaminhan-Hassemer M, Michelato M, Boscolo WR et al. Dietary histidine requirement of grow-out Nile tilapia (*Oreochromis niloticus*), based on growth performance, muscle development, expression of muscle growth-related genes, and blood parameters. Revista Brasileira de Zootecnia, 2020;49 e20180210.
73. Breck O, Bjerkas E, Campbell P et al. Histidine nutrition and genotype affect cataract development in Atlantic salmon, *Salmo salar* L. Journal of Fish Diseases, 2005;28(6), 357-371. Doi: 10.1.111/j.1365-2761.2.005.0.0640.x
74. Remo SC, Hevroy EM, Olsvik PA et al. Dietary histidine requirement to reduce the risk and severity of cataracts is higher than the requirement for growth in Atlantic salmon smolts, independently of the dietary lipid source. British Journal of Nutrition, 2014;111(10), 1759-1772. DOI: 10.1.017/S0007114513004418
75. Steinberg CE. Aquatic animal nutrition. 2022. Springer International Publishing.
76. Sohn KS, Kim MK, Kim JD, Han IK. The role of immunostimulants in monogastric animal and fish-review. Asian-Australasian Journal of Animal Sciences, 2000;13(8), 1178-1187.
77. Hernandez LH, Hardy RW. Vitamin A functions and requirements in fish. Aquaculture Research, 2020;51(8), 3061-3071.
78. Vijayan KK, Makesh M, Otta SK, et al. Prophylaxis in aquaculture. 2017; Chennai, India: ICAR. Obtenido de <https://krishi.icar.gov.in/jspui/bitstream/123456789/10377/1/Compendium%20on%20Prophylaxis%20in%20Aquaculture.pdf>.
79. Cao ZH, Luo JB, Yu WB. Effect of vitamin C on growth and non-specific immune of finless eel (*Monopterus albus*). Journal of Yangtze University 2009;6, 27-29
80. Lock EJ, Waagbø R, Wendelaar Bonga S, Flik G. The significance of vitamin D for fish: a review. Aquaculture nutrition, 2010;16(1), 100-116.
81. Cheng K, Huan Y, Wang C et al. Physiological function of vitamin D3 in fish. Reviews in Aquaculture, 2023;15(4), 1732-1748.
82. Liu A, Santigosa E, Dumas A, Hernandez JM. Vitamin nutrition in salmonid aquaculture: From avoiding deficiencies to enhancing functionalities. Aquaculture, 2022;561, 738654.
83. Ferencik M, Ebringer L. Modulatory effects of selenium and zinc on the immune system. Folia microbiologica, 2003;48, 417-426.
84. Musharraf M, Khan MA. Dietary zinc requirement of fingerling Indian major carp, *Labeo rohita* (Hamilton). Aquaculture, 2019;503, 489-498.
85. Dawood MA, Basuini MFE, Yilmaz S et al. Selenium nanoparticles as a natural antioxidant and metabolic regulator in aquaculture: A review. Antioxidants, 2021;10(9), 1364. DOI:10.3390/antiox10091364n, 2021).
86. Wangkahart E, Bruneel B, Chantiratikul A et al. Optimum dietary sources and levels of selenium improves growth, antioxidant status, and disease resistance: Re-evaluation in a farmed fish species, Nile tilapia (*Oreochromis niloticus*). Fish & Shellfish Immunology. 2021. DOI:10.1016/j.fsi.2021.12.003
87. Hermes LB, Peixoto NC, Battisti EK et al. Dietary iron affect innate immunity, hematologi-

- cal and oxidative responses in Nile tilapia (*Oreochromis niloticus*). *Aquaculture International*, 2024;32(4), 3993-4007.
88. Evliyaoğlu E, Kilercioğlu S, Yılmaz HA, et al. Iron supplementation in plant-based aquafeed: Effects on growth performance, tissue composition, iron-related serum parameters and gene expression in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*, 2022;550, 737884.
89. Ibrahim D, Rahman MIA, Abd El-Ghany AM et al. *Chlorella vulgaris* extract conjugated magnetic iron nanoparticles in Nile tilapia (*Oreochromis niloticus*): Growth promoting, immunostimulant and antioxidant role and combating against the synergistic infection with *Ichthyophthirius multifiliis* and *Aeromonas hydrophila*. *Fish & Shellfish Immunology*, 2024;145, 109352